Blood components used for transfusion therapy include platelet concentrates (PCs), red blood cell concentrates (RBCC), and cell-free plasma. Significant advances have been made to increase the safety of these blood products in recent years by reducing the occurrence of units contaminated with viruses such as HIV. However, bacterial contamination of PCs, used as a therapeutic product to treat thrombocytopenic or bleeding patients, has become the most significant post-transfusion infectious risk in developed countries (1,2). PCs are exquisitely susceptible to bacterial proliferation, in comparison to RBCC and plasma, due to their storage conditions in gas-permeable plastic containers, at temperatures of 20–24 ℃, under agitation for up to 7 days. PCs are prepared in either 100% plasma or in platelet additive solutions (PAS) containing high glucose concentration. These storage conditions are important to maintain platelet functionality; however, they also provide an ideal environment for bacterial propagation. The predominant bacteria isolated from contaminated PCs are Gram-positive organisms, which are part of the normal skin or mucosa flora of the blood donor (1,2). Less frequently, Gram-negative organisms, which may be part of transient skin flora or originate from silent donor bacteremia, are isolated from contaminated PCs. Usually, transfusion reactions involving Gram-negative bacteria are more severe due to infused endotoxin (lipopolysaccharide of the cell wall) followed by massive cytokine release. Clinical symptoms may include fever over 38.5 ℃, hypotension, nausea, vomiting and septic shock (2).

Although bacterial contamination is usually originated from the donor, there is also a potential for PC contamination during blood collection, storage, or even retrograde contamination from the patient to the PC component (3-5). Measures implemented worldwide to mitigate the risk of transfusing bacterially-contaminated PCs include donor screening with a questionnaire, skin disinfection of the donor’s venipuncture site, diverting the first aliquot (approximately 30–40 mL) of the donated blood, PC screening for the presence of bacteria with culture or rapid methods, and pathogen reduction technologies (PRT) (1,2).

In the February 2020 issue of Transfusion (6), Emery et al. reported a genotypic study of five *Citrobacter koseri* strains isolated from contaminated PCs in France from 2012 to 2017. The Gram-negative bacillus *C. koseri* is part of the normal flora of human and animal digestive systems. Two of the isolates were traced back to the donors. Strain PAR was isolated from the donor’s nose and strain NAN was retrieved from the donor’s armpit. It is therefore likely that *C. koseri* can transiently colonize human skin and mucosa. Three of the five *C. koseri* strains described in this study were implicated in septic transfusion events with two resulting in fatalities (7,8). The other two strains were isolated from contaminated PCs that were discarded prior to transfusion into patients. The authors conducted comprehensive phylogenetic analyses of the five PC...
C. koseri isolates in comparison to five C. koseri strains recovered from human samples but unrelated to transfusion events. The genomes of the strains were sequenced and assembled along fifteen C. koseri genomes available in NCBI databases. A genome multilocus sequence typing (cgMLST) scheme was constructed. The genomic comparison identified 4,950 genes shared by ≥96% of the selected genomes. This approach was used to visualize evolutionary relationships within the C. koseri isolates. Results of the genomic study showed that the PC isolates were nonclonal and did not share specific genes. However, one cluster of 11 strains, including three of the five PC isolates (BES, PAR and NAN), was identified. The origin of the 11 strains is distributed worldwide and therefore a potential common origin is unlikely. The authors also tested the growth characteristics of the PC isolates in PCs prepared in plasma or PAS and found no differences in growth characteristics.

Clusters of transfusion-associated septic reactions involving bacterially-contaminated PCs have been reported in the past. In 1991, six patients in Denmark and Sweden developed septicemia with Serratia marcescens after PC transfusion (3). The source of contamination was found on the exterior of blood bags produced in Belgium. In 2017, two separate clusters of fatal septic transfusion reactions involving PCs contaminated with Clostridium perfringens and Klebsiella pneumoniae were documented in the US, in Utah and California, respectively (9). More recently, multiple septic reactions with a potential common source were reported in three US states involving PCs contaminated with Acinetobacter calcoaceticus-baumannii and Staphylococcus saprophyticus (4). Investigation of these cluster cases with molecular testing of isolates from the donors, PC bags and platelet incubators concluded that a potential common source of contamination was likely responsible for the multiple septic transfusion cases. Unfortunately, in the C. koseri outbreak case discussed herein, there was not microbiological investigation of the PAS used for PC manufacturing, or the equipment and materials used during PC collection, production, or storage. Such investigation would have been especially relevant to further study the relationship of C. koseri strains BES and NAN, which were both isolated in 2017 from whole-blood derived PCs. These two strains belong to the cluster identified during cgMLST. Although they were isolated in different French cities, the possibility of a common source of contamination covering different geographic locations exists as demonstrated in the Acinetobacter calcoaceticus-baumannii and Staphylococcus saprophyticus case mentioned above (4).

Definite and probable published transfusion septic cases of PCs contaminated with C. koseri are summarized in Table 1. In addition to the three PC contamination cases reported by Emery et al. (6), there was another report of a septic transfusion case involving PCs contaminated with C. koseri, also in France, in 2015 (11). It is interesting that these reports are concentrated in France during the period of 2012 to 2017. C. koseri was first described to be involved in a septic transfusion event implicating contaminated RBCC in Brazil, in 2012 (10). The Japanese Red Cross also documented a severe septic transfusion case with PCs contaminated with C. koseri in 2016 (12). The Japanese report emphasized the importance of visual inspection prior to transfusion of PCs. C. koseri induces platelet clump formation when grown in PCs as shown in bulletins of the Japanese Red Cross and the Australian Red Cross Blood Service (12,13). Platelet aggregation in contaminated PCs is a common feature triggered by other species including Staphylococcus aureus, Klebsiella pneumoniae, and Enterobacter aerogenes (12,14-16).

As stated by the authors, in France, blood products suspected of being contaminated with bacteria are

<table>
<thead>
<tr>
<th>Year of transfusion event</th>
<th>Origin</th>
<th>Blood component</th>
<th>Clinical outcome</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>2008</td>
<td>Brazil</td>
<td>RBCC</td>
<td>Severe septic reaction</td>
<td>(10)</td>
</tr>
<tr>
<td>2012</td>
<td>France</td>
<td>Apheresis PCs</td>
<td>Septic reaction</td>
<td>(6)</td>
</tr>
<tr>
<td>2015</td>
<td>France</td>
<td>Apheresis PCs</td>
<td>Fatality</td>
<td>(6,7)</td>
</tr>
<tr>
<td>Unknown</td>
<td>France</td>
<td>PCs</td>
<td>Septic shock</td>
<td>(11)</td>
</tr>
<tr>
<td>2017</td>
<td>France</td>
<td>Whole-blood-derived PCs</td>
<td>Septic shock</td>
<td>(6,8)</td>
</tr>
<tr>
<td>2016</td>
<td>Japan</td>
<td>Apheresis PCs</td>
<td>Severe septic reaction. Patient recovered with sequelae</td>
<td>(12)</td>
</tr>
</tbody>
</table>

RBCC, red blood cell concentrates; PCs, platelet concentrates.

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Ann Blood 2020 | http://dx.doi.org/10.21037/aob-20-41
quarantined and discarded, and definite or probable transfusion-transmitted cases are reported to the French hemovigilance organization. Several mitigation strategies to prevent transfusion of contaminated PC units have been implemented in France although PC screening is not one of them. Automated culture systems are effective in capturing contaminated PCs with Gram-negative bacteria which in general proliferate fast in this blood component (17,18). As shown in the supplemental material, of the Emery et al. publication (6), C. koseri reaches concentrations >10^3 colony forming units (CFU)/mL after 24 hours of PC storage. These results indicate that C. koseri grows fast in PCs and could be captured by automated culture systems preventing transfusion of contaminated PCs with this bacterium as reported by Canadian Blood Services (17). Although there are no published reports of inactivation of C. koseri with PRT, it is important to note that the PRT Intercept™ (CERUS Corp.) was implemented in France in 2017. Since then, there have not been reports of septic transfusion reactions involving PCs contaminated with C. koseri. It is however imperative to consider the potential of bacterial contamination post-PR treatment during PC storage or transportation as discussed by Jones et al. (4).

Overall, bacterial contamination of PCs poses the most prevalent transfusion-transmitted infectious risk due to their storage conditions. Several interventions have decreased but not eliminated the occurrence of transfusion-associated septic events involving contaminated PCs. Although Gram-positive skin/mucosa flora are the predominant PC contaminants, Gram-negative bacteria, such as C. koseri, can contaminate PCs posing a major infectious risk due to endotoxin release, resulting in septic shock in transfusion patients. A cluster of five PCs contaminated with C. koseri documented in France from 2012 to 2017 has been discussed herein. Importantly, no more septic cases involving PCs contaminated with this organism have been reported since the implementation of PRT in France in 2017. Outbreaks of transfusion septic events require through investigations of the PC donor, PC recipient, and equipment involved during blood collection, and PC production and storage. Recognizing, investigating, and reporting septic transfusion reactions to hemovigilance systems is highly recommended to interdict the transfusion of contaminated PC units.

Acknowledgments

Funding: Funding for this study was provided by Canadian Blood Services and Health Canada. The views expressed herein do not necessarily represent the view of the federal government.

Footnote

Conflicts of Interest: The author has completed the ICMJE uniform disclosure form (available at http://dx.doi.org/10.21037/aob-20-41). The author has no conflicts of interest to declare.

Ethical Statement: The author is accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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Cite this article as: Ramirez-Arcos S. Transfusion septic reactions involving platelet concentrates contaminated with Citrobacter koseri. Ann Blood 2020.