

# Incidence and severity of adverse effects related to platelet transfusion: a narrative review of the literature and the recent hemovigilance data of Japan

Tomohiko Sato<sup>1</sup>, Nelson Hirokazu Tsuno<sup>2</sup>, Naoko Goto<sup>3</sup>, Takeshi Hagino<sup>4</sup>, Tetsunori Tasaki<sup>1</sup>

<sup>1</sup>Department of Transfusion Medicine and Cell Therapy, The Jikei University Hospital, Minato-ku, Tokyo, Japan; <sup>2</sup>Japanese Red Cross Society Kanto-Koshientu Block Blood Center, Koto-ku, Tokyo, Japan; <sup>3</sup>Safety Vigilance Division, Technical Department, Blood Service Headquarters, Japanese Red Cross Society, Minato-ku, Tokyo, Japan; <sup>4</sup>Department of Hematology, Tama-Hokubu Medical Center, Tokyo Metropolitan Health and Medical Treatment Corporation, Higashimurayama, Tokyo, Japan

**Contributions:** (I) Conception and design: T Sato, NH Tsuno, T Hagino, T Tasaki; (II) Administrative support: T Sato, NH Tsuno, T Tasaki; (III) Provision of study materials or patients: NH Tsuno, N Goto; (IV) Collection and assembly of data: T Sato, NH Tsuno, N Goto, T Hagino; (V) Data analysis and interpretation: All authors; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

**Correspondence to:** Tomohiko Sato. Department of Transfusion Medicine and Cell Therapy, The Jikei University Hospital, 3-19-18, Nishi-Shimbashi, Minato-ku, Tokyo, 105-8471, Japan. Email: tomosatou@jikei.ac.jp.

**Abstract:** Platelet transfusions are given to prevent or treat bleeding in patients with quantitative or qualitative platelet disorders. Despite these lifesaving roles, certain complications are unavoidable even with best practices. Administration of platelet components has various risks, including allergic and anaphylactic transfusion reactions, febrile non-hemolytic transfusion reactions (FNHTRs), bacterial and viral infections, transfusion-related acute lung injury (TRALI), transfusion-associated circulatory overload (TACO), transfusion-associated graft-versus-host disease (TA-GVHD) etc., some of which can be fatal. Among them, allergic and anaphylactic transfusion reactions, FNHTRs are frequent with platelet and plasma transfusions. Due to storage at room temperature, bacterial infections are more frequently caused by platelet transfusions than red blood cell or plasma transfusions. To prevent these adverse events related to platelet transfusions, various safety measures are being applied, which include well-organized donor-selection protocol, skin disinfection, sample diversion, leukoreduction, detection of bacterial contamination by culture and/or immunoassay, plasma replacement with platelet additive solution, platelet washing, and pathogen reduction technology. In contrast to other developed countries, the situation of platelet transfusions in Japan is unique in that the shelf-life of leukoreduced single-donor apheresis platelets is limited to 3 days and bacterial culture screening and pathogen reduction technologies are not implemented so far. In this article, incidence and severity of various adverse effects of platelet transfusions in Japan are described by referring to the recent data collected by two existing hemovigilance systems at the national level. Moreover, existing literatures inside and outside Japan regarding epidemiology, clinical manifestation, treatment, and mitigation strategies of adverse events associated with platelet transfusions are reviewed, which can help clinicians' appropriate use of platelet components and management of related adverse events.

**Keywords:** Platelet transfusion; adverse transfusion reaction; transfusion-transmitted infection (TTI); hemovigilance

Received: 16 November 2020; Accepted: 28 December 2020; Published: 25 September 2021.

doi: 10.21037/aob-20-90

**View this article at:** <http://dx.doi.org/10.21037/aob-20-90>

## Introduction

In clinical practice, platelet transfusions are indicated for prevention or treatment of bleeding in patients with either low platelet counts or poor platelet function (1-3), the majority of which comprise of prophylactic transfusion for patients with chemotherapy or hematopoietic progenitor cell transplantation to reduce the risk of spontaneous bleeding (4-7). Other cases, mostly in general medicine, cardiac surgery, intensive care unit etc., require therapeutic platelet transfusions for treating acute hemorrhage (6,7). Platelets can be collected by isolating and pooling platelets from units of donated whole blood (pooled whole-blood-derived platelets) or by apheresis directly from a donor (single-donor apheresis platelets) (1,8). Compared to red blood cells (RBCs) and plasma, which are stored at 4 °C for 5 to 6 weeks and at -20 °C for about 1 year, respectively, platelets are stored at room temperature (20 to 24 °C) in gas permeable bags with constant agitation to preserve the viability. Thus, in most of developed countries, platelet shelf-life is limited to no longer than 5 days because of the bacterial infection risk that increases in relationship to the storage duration (9,10).

Transfusion practice is inseparable from adverse events, which include adverse transfusion reactions and transfusion-transmitted infections (TTIs). Incidence of the frequent adverse reactions to any blood components is estimated to be about one in 100 transfusions, and some reactions can be fatal, although very rare, occurring in about one in 200,000 to 420,000 units (11,12). Incidence of adverse reactions associated with platelet transfusions is approximately 1% to 4%, which is higher compared with those with RBC or plasma transfusions (13-16). According to the definition of adverse transfusion reactions by the International Society of Blood Transfusion (ISBT) Working Party for Hemovigilance (17), most of reactions related to administration of platelet concentrate (PC) are classified as either allergic transfusion reactions (ATRs) or febrile non-hemolytic transfusion reactions (FNHTRs) (16,18). These major reactions have been shown to be caused by the supernatant plasma in which units are stored (19,20). As for TTIs, bacterial infections are relatively more common with platelet transfusions than with RBC or plasma transfusions due to storage of platelets at warmer temperature.

Leukoreduction is a major safety measure to prevent adverse events related to platelet transfusions that has been widely implemented in developed countries, which is beneficial for reducing FNHTRs, bacterial and viral

infections (12,21-23). Replacement of plasma in PC bags with platelet additive solution (PAS) and platelet washing are safety measures to prevent ATRs (23-27). The common preventive strategies for transfusion-transmitted bacterial infection (TTBI) include well-organized donor-selection protocol, skin disinfection, sample diversion, leukoreduction, and detection of contamination by culture and/or immunoassay (12,23,28-32). Pathogen reduction technology (PRT) has been proven to reduce the residual risk of TTIs (8,12,28,33,34). There is an inter-country variation in adopting safety measures for preventing adverse events of platelet transfusions, and the situation in Japan is unique (35-37). However, information regarding platelet transfusion in Japan has been poorly described in existing literatures.

This article aims at reviewing existing literatures inside and outside Japan regarding epidemiology, clinical manifestation, treatment, and mitigation strategies of adverse events associated with platelet transfusions. Additionally, this article provides incidence and severity of these events in Japan using the recent data collected by two existing hemovigilance systems at the national level. This article is presented in accordance with the narrative review checklist (available at <http://dx.doi.org/10.21037/aob-20-90>).

## Literature search strategy

The medical literatures for published observational/ investigational studies, randomized controlled trials or systematic reviews and meta-analyses regarding adverse events related to platelet transfusions were analyzed. The PubMed/Medline electronic database was searched in November 2020 using the primary phrases such as “platelet transfusion”, “adverse transfusion reaction”, “transfusion-transmitted infection”, “incidence”, “severity”. Further search was performed with additional key terms such as “allergic transfusion reaction”, “febrile non-hemolytic transfusion reaction”, “anaphylactic transfusion reaction”, “bacterial infection”, “mitigation strategy”, “safety measure”. Moreover, a hand search of *ISBT Science Series* and *Annals of Blood* was also added. English language, peer-reviewed articles were considered and no constraints on publication type or date were imposed. Titles/abstracts of retrieved articles were checked for relevance, and other relevant papers were identified by manual searching of reference lists and the authors' personal literature collections. When similar findings were reported in

**Table 1** Comparisons of TTBI mitigation strategies and PC shelf-life

Area	Country	Shelf-life, days	Bacterial screening, implementation	Pathogen inactivation, implementation	Fatal TTBI, cases/1M bags	Ref.
Europe	Belgium	7	+	Universal	0/0*	(22,46)
	France	7	–	Universal	3.5/0*	(22,46)
	Switzerland	5	–	Universal	19.1/0*	(22,46)
	United Kingdom	7	+	–	2.3	(22,46)
America	United States	5	+	Available	1.3	(43)**
	Canada	7	+	Available	2	(44,47)
Asia/ Pacific	Australia	5	+	Available	0	(29)
	New Zealand	7	+	–	0	(45)
	Japan	3	–	–	0.1	(36)***

\*, before/after implementation of pathogen reduction technology; \*\*, can be extended to 7 days if negative by rapid immunoassay; \*\*\*, all single-donor apheresis platelets are inspected visually. TTBI, transfusion-transmitted bacterial infection; PC, platelet concentrate.

multiple articles, priority was given to those most recently published.

### Production and supply of PCs in Japan

The Japanese Red Cross Society (JRCS), as a sole operator of blood services, controls blood collection, processing and supply of blood products nationwide (38,39). In Japan, all PCs transfused are leukoreduced single-donor apheresis platelets. Of 4,707,951 voluntary non-remunerated donations in 2018, platelet apheresis donations accounted for 620,414 (13.2%), and 808,179 PC bags were distributed nationwide (40,41).

Eligibility criteria for platelet apheresis donation are as follows: age of 18 to 69 for male, 18 to 54 for female, body weight of 45 kg or more for male and 40 kg or more for female, hemoglobin level of 12 g/dL or more, and platelet count from 0.15 to 0.6 million/ $\mu$ L. After donor's skin disinfection with 10% povidone-iodine followed by 70% isopropyl alcohol and the diversion of the initial 25 mL portion of blood, up to 600 mL of platelets suspended in plasma can be collected by any of three collection systems: CCS (Haemonetics), Terusys-S (Terumo), and Trima (Terumo) (36,39). More than 0.2, 0.4, 1.0, 2.0, 3.0, 4.0  $\times 10^{11}$  platelets are contained in 1-, 2-, 5-, 10-, 15-, 20-unit PC bags respectively, and 10-unit PC bags (volume of about 200 mL) are the most commonly used in clinical practice (35). All PCs are leukoreduced before storage, stored at 20 to 24 °C with agitation, and visually inspected

for the presence of aggregates and/or swirling before issue (36).

ABO-identical, single-donor apheresis platelets are routinely supplied by the JRCS, however, human leukocyte antigen (HLA) and/or human platelet antigen (HPA) matching is given priority over ABO compatibility when there is a need for HLA- or HPA-matched platelets. Currently, approximately 70% of supplied HLA-matched platelets are ABO-identical. Non-ABO-identical HLA- or HPA-matched platelets with high ABO antibody titers ( $>1:128$ ) are administered at the physician's discretion (42).

The JRCS has already implemented universal leukoreduction and initial flow diversion, but not bacterial culture screening or PRTs so far (36). For reducing clinically relevant risk of sepsis due to TTBI, the shelf-life of PC is presently limited to 3 days. This is the shortest when compared to other developed countries (22,29,43–45), where bacterial screening of PC bags and/or PRTs are being implemented as strategies to mitigate TTBI (Table 1). This “3-day limitation policy” has contributed to the low incidence of fatal TTBI related to platelet transfusion in Japan, which is currently estimated to be 0.1 in 1 million PC bags (36).

Reflecting effectiveness of decreasing the volume of plasma for preventing ATRs, which contains inflammatory cytokines and protein components (48–50), the JRCS started to provide washed PCs in 2016 (51). In contrast to Western countries, PAS-PCs, in which about 70% of plasma is replaced with PAS, are not available so far in Japan.

**Table 2** Hemovigilance data by the JRCS during 2010–2018: mean frequencies of adverse reactions

Adverse reactions	PC		RBC		FFP	
	Mean	95% CI	Mean	95% CI	Mean	95% CI
Allergic reaction	35.86	22.31–49.41	4.45	3.62–5.29	11.97	9.68–14.27
Severe allergic reaction	25.02	22.29–27.73	2.72	2.04–3.41	8.73	8.19–9.27
Fever	5.58	4.37–6.79	3.66	2.85–4.47	0.62	1.06–1.48
Respiratory distress	5.16	3.915–6.399	2.42	2.03–2.80	1.21	0.91–1.52
Hypotension	2.59	2.214–2.970	1.37	1.17–1.57	1.27	0.33–0.91
TACO	0.48	0.203–0.749	0.58	0.33–0.83	0.22	0.08–0.36
TRALI	0.36	0.225–0.503	0.12	0.04–0.17	0.14	0.05–0.22
Others	3.81	2.649–4.974	2.09	1.53–2.65	0.62	0.42–0.82
Total	78.98	65.32–92.62	17.52	15.51–19.53	24.83	22.40–27.25
No. of bags distributed in Japan*	819,691	808,885–830,497	3,364,908	3,310,163–3,419,653	961,589	945,373–977,804

Mean frequencies of adverse events per 100,000 transfusion bags. \*, derived from hemovigilance data by the JRCS (35). JRCS, Japanese Red Cross Society; PC, platelet concentrate; RBC, red blood cell; FFP, fresh frozen plasma; CI, confidence interval; TACO, transfusion-associated circulatory overload; TRALI, transfusion-related acute lung injury.

## Recent hemovigilance data of Japan

### *Two existing hemovigilance systems*

Presently, two hemovigilance data are available in Japan, one gathered by the JRCS (40), and the other by the National Institute of Infectious Diseases (NIID) of Japan and the Hemovigilance Committee of the Japan Society of Transfusion Medicine and Cell Therapy (JSTMCT). The JRCS established a hemovigilance system in 1993, in accordance with Pharmaceuticals and Medical Devices act of Japan. The system can provide surveillance of transfusion adverse reactions and infections nationwide (52), with voluntary paper-based reports from domestic hospitals and information from blood donors. Since that, approximately 2,000 suspected cases with moderate to severe adverse events have been reported annually (40), however, the largest and voluntary reporting system has precluded surveillance of mild adverse events (38).

Thereafter, to collect comprehensive data on adverse transfusion reactions nationwide, an online reporting system for transfusion-related adverse events was introduced in 2007 by the NIID and the JSTMCT (38,51). With voluntary participation of academic hospitals and small-scale hospitals, the incidence of adverse events to PCs recorded in the online system was shown to be comparable to the rates reported in other countries (38,51). The surveillance by the online reporting system has been so far monitoring

about 10% of the bags distributed in Japan (53).

The hemovigilance situations on monitoring adverse events to blood transfusion vary from country to country. It is mandatory to report fatal cases in the US (54), serious or unexpected adverse reactions in Canada (44), serious adverse reactions and events in the UK and Belgium (55,56), any suspected transfusion reactions in France and Switzerland (57,58). On the other hand, in Australia and New Zealand, reporting of serious adverse reactions is performed on a voluntary basis, as is in Japan (45,59).

Hereafter in this article, the online hemovigilance data by the NIID (53) from 2010 to 2018 was evaluated mainly in terms of mild adverse events associated with platelet transfusions. During this period, approximately 40 to 50 hospitals per year {academic hospitals: median 43 [34–46]; small-scale hospitals: median 4 [3–5]} participated in the online surveillance by the NIID. The paper-based data by the JRCS (40) from 2010 to 2018 was used to evaluate mainly severe adverse events.

### *Frequent adverse events related to platelet transfusions in Japan*

Table 2 shows the incidence of adverse reactions reported to the JRCS from 2010 to 2018. The mean numbers of PC, RBC, and fresh frozen plasma (FFP) bags distributed yearly in Japan were 819,691 (95% CI: 808,885–830,497),

**Table 3** Hemovigilance data by the NIID during 2010–2018: clinical diagnoses of adverse events

Clinical diagnoses	PC		RBC		FFP	
	Mean	95% CI	Mean	95% CI	Mean	95% CI
Non-hemolytic transfusion reactions*	3,156.93	2,810.37–3,503.48	636.32	589.38–683.26	1,117.35	971.58–1,263.12
Other non-hemolytic reaction	3,120.57	2,780.86–3,460.28	627.81	580.85–674.77	1,095.87	952.15–1,239.58
Severe allergic reaction	33.93	22.53–45.34	14.23	–1.02 to 29.48	19.10	14.36–23.84
TRALI	1.53	0.78–2.28	0.62	0.26–0.98	1.48	0.14–2.82
TACO	0.89	0.27–1.52	1.29	0.95–1.62	0.91	0.29–1.53
GVHD	0.00	–	0.00	–	0.00	–
PTP	0.00	–	0.00	–	0.00	–
Hemolytic reaction*	0.00	–	1.29	0.46–2.11	0.00	–
Acute hemolytic reaction	0.00	–	0.67	0.18–1.17	0.00	–
Delayed hemolytic reaction	0.00	–	0.62	0.13–1.10	0.00	–
Infectious diseases*	0.51	–0.03 to 1.06	0.06	–0.05 to 0.17	0.23	–0.07 to 0.52
Bacterial infection	0.38	0.00–0.76	0.00	–	0.00	–
HBV infection	0.13	–0.12 to 0.38	0.00	–	0.23	–0.07 to 0.52
HCV infection	0.00	–	0.00	–	0.00	–
HIV infection	0.00	–	0.00	–	0.00	–
Other infections	0.00	–	0.06	–0.05 to 0.17	0.00	–
Total*	3,157.44	2,811.05–3,503.82	637.67	591.04–684.29	1,117.58	971.92–1,263.24
No. of bags reported to the NIID	109,389	100,965–117,812	242,846	223,527–262,164	124,567	112,644–136,490
% of bags reported/distributed	13.35	12.39–14.27	7.22	6.69–7.73	12.95	11.82–14.05
No. of bags distributed in Japan**	819,691	808,885–830,497	3,364,908	3,310,163–3,419,653	961,589	945,373–977,804

\*, mean frequencies of adverse events per 100,000 transfusion bags; \*\*, derived from hemovigilance data by the JRCS (35). NIID, National Institute of Infectious Diseases; PC, platelet concentrate; RBC, red blood cell; FFP, fresh frozen plasma; CI, confidence interval; TRALI, transfusion-related acute lung injury; TACO, transfusion-associated circulatory overload; GVHD, graft-versus-host disease; PTP, post-transfusion purpura.

3,364,908 (95% CI: 3,310,163–3,419,653), and 961,589 (95% CI: 945,373–977,804), respectively. The mean incidences of all adverse reactions associated with PC, RBC, and FFP were 78.98 (95% CI: 65.32–92.62), 17.52 (95% CI: 15.51–19.53), and 24.83 (95% CI: 22.40–27.25) per 100,000 distributed bags, respectively. Thus, according to the JRCS data, ATRs were the most frequent adverse events of platelet transfusions, with an incidence of 0.03% (35.86 per 100,000 bags).

Table 3 shows the clinical diagnoses of adverse events

reported online to the NIID from 2010 to 2018. The mean numbers of PC, RBC, and FFP bags transfused at participating hospitals were 109,389 (95% CI: 100,965–117,812), 242,846 (95% CI: 223,527–262,164), and 124,567 (95% CI: 112,644–136,490), respectively, accounting for 13.35%, 7.22%, and 12.95% of bags distributed in Japan, respectively. The mean incidences of all adverse reactions associated with PC, RBC, and FFP were 3,157.44 (95% CI: 2,811.05–3,503.82), 637.67 (95% CI: 591.04–684.29), and 1,117.58 (95% CI: 971.92–1,263.24) per 100,000

**Table 4** Hemovigilance data by the NIID during 2010–2018: clinical signs of adverse events

Clinical signs	PC		RBC		FFP	
	Mean	95% CI	Mean	95% CI	Mean	95% CI
Urticaria	2,339.95	2,066.45–2,613.44	215.97	194.20–237.74	921.92	782.78–1,061.06
Pruritus	1,336.07	1,178.77–1,493.37	78.41	71.36–85.46	446.46	363.87–529.06
Rash	392.67	345.56–439.77	66.37	60.19–72.55	220.45	178.05–262.85
Fever	278.49	242.86–314.11	201.07	184.91–217.24	71.63	57.11–86.14
Feverishness	105.38	95.54–115.20	54.83	48.81–60.86	52.18	41.72–62.65
Chill, rigor	100.02	88.99–111.04	40.44	35.23–41.64	37.97	28.18–47.77
Respiratory distress	87.64	74.62–100.65	22.74	20.25–25.23	40.59	32.93–48.25
Hypotension	64.55	55.02–74.08	34.05	29.00–39.10	66.05	56.27–75.84
Nausea, vomiting	50.65	43.38–57.90	29.01	25.36–32.66	34.90	24.01–45.80
Hypertension	41.08	32.72–49.42	49.23	41.28–57.18	16.03	11.18–20.89
Tachycardia	35.85	30.15–41.54	16.97	14.76–19.19	20.24	17.29–23.18
Chest, flank or back pain	15.18	12.61–17.74	9.41	6.67–12.15	6.48	4.26–8.70
Headache	14.54	10.96–18.12	9.24	7.22–11.26	4.43	2.48–6.38
Consciousness disturbance	5.49	3.58–7.38	0.95	0.26–1.64	0.45	–0.03 to 0.94
Venous pain	2.93	1.62–4.23	14.56	13.00–16.13	0.34	–0.13 to 0.81
Hematuria	1.15	0.61–1.67	1.68	0.73–2.63	0.57	0.08–1.05
Others	42.23	31.50–52.95	15.85	10.58–21.13	27.06	15.45–38.67
Total	4,913.86	4,381.31–5,446.40	860.81	802.83–918.78	1967.76	1,669.63–2,265.89

Mean frequencies of adverse events per 100,000 transfusion bags. NIID, National Institute of Infectious Diseases; PC, platelet concentrate; RBC, red blood cell; FFP, fresh frozen plasma; CI, confidence interval.

bags, respectively. Thus, according to the NIID data, non-hemolytic reactions, mostly ATRs and FNHTRs, were the most frequent adverse events of platelet transfusions, with an incidence of 3.16% (3,156.93 per 100,000 bags).

Table 4 shows the clinical signs of adverse events reported online to the NIID from 2010 to 2018. Urticaria, pruritus, rash, fever, and feverishness were the most frequently reported signs, with incidences of 2,339.95, 1,336.07, 392.67, 278.49, and 105.38 per 100,000 bags, respectively. ATR-related signs (urticaria, pruritus, rash) represented about 80% of all reported adverse events.

This big difference in the incidence of mild adverse reactions, such as ATRs and FNHTRs, between the JRCS and the NIID data might be dependent on the different scopes of each reporting system. Since most clinicians report suspected cases of adverse events to the JRCS with the intent to have the causal relationship investigated, the

reported cases tend to be of moderate to severe intensities, and mild cases are usually underreported. Reflecting the nationwide coverage of the JRCS reporting system, it is suitable for the analysis of moderate to severe adverse events in Japan. On the other hand, mild adverse reactions are better collected by the online reporting system of the NIID, through passive surveillance (60), where the blood users of the participating hospitals register the whole hemovigilance data (38).

### *Severe adverse events related to platelet transfusions in Japan*

Table 5 shows the severe adverse events related to all blood components reported to the JRCS from 2010 to 2018, which included fatal cases and non-fatal cases that required intensive treatment. The numbers of cases with TTI, severe



**Table 5** Severe adverse events reported to the JRCS during 2010–2018

Year	2010	2011	2012	2013	2014	2015	2016	2017	2018	Total
Severe allergic reaction	326	353	357	280	464	497	405	360	347	3,389
TRALI*	9 [2]	14	6	9	2 [1]	7	4	5	4	60 [3]
p-TRALI*	15 [1]	10	4	10	7	6	3	4	1	60 [1]
TACO	0	0	26	29	44	63	45	47	55	309
Cardiogenic pulmonary edema	47	48	44	35	33	38	40	35	49	369
Respiratory distress total*	71 [3]	72	80	83	86 [1]	114	92	91	109	798 [4]
Bacteria infection										
Suspicious	28	21	31	25	17	25	20	21	23	211
Confirmed*	0	1	1	1	0	2	1	3 [1]	4	13 [1]
HBV infection										
Suspicious	36	40	50	52	27	14	18	20	13	270
Confirmed*	11	13	6	7	2	0	1	1	1	42
HCV infection										
Suspicious	27	29	40	32	26	35	28	21	17	255
Confirmed*	2	0	0	1	0	0	0	0	0	3
Other infections										
Suspicious	7	6	10	16	11	19	14	12	19	114
Confirmed*	1	1	4	2	4	4	3	4 [1]	7	30 [1]
Infection total										
Suspicious	98	96	131	125	81	93	80	74	72	850
Confirmed*	14	15	11	11	6	6	5	8 [2]	12	88 [2]
No. of transfusion bags distributed**	5,197,025	5,237,637	5,287,450	5,243,088	5,160,350	5,103,352	5,060,832	5,041,752	4,984,201	46,315,687

\*, no. of confirmed cases [No. of fatal cases]; \*\*, total no. of distributed bags including PC, RBC, and FFP, derived from hemovigilance data by the JRCS (35). JRCS, Japanese Red Cross Society; TRALI, transfusion-related acute lung injury; p-TRALI, possible TRALI; TACO, transfusion-associated circulatory overload; HBV, hepatitis B virus; HCV, hepatitis C virus.

allergic reaction, and respiratory distress were 88, 3,389, and 798, respectively, with the incidence of 0.17, 7.31, and 1.38 per 100,000 transfusion bags (including PC, RBC, and FFP). During the period, a total of six fatal cases were reported, which included one case of TTBI (*E. coli*), one case of viral infection [hepatitis E virus (HEV)], three cases of TRALI, and one case of possible TRALI (p-TRALI). Data regarding fatal TACO cases was unavailable in this period mainly because TACO reporting criteria of the surveillance by the JRCS were under consideration.

### Management of frequent adverse events related to platelet transfusions

ATRs and FNHTRs are frequent adverse reactions associated with platelet transfusions, and these incidences are reported to be higher than those associated with RBC or plasma transfusions (16,18,61,62).

#### ATR

ATRs are the most frequent adverse events of platelet

transfusions (62), with a reported incidence of 3% to 7% (12,63). Typical symptoms such as urticaria, rash, pruritus, and localized angioedema are usually mild, however, anaphylaxis, the severe form of ATRs, sometimes occur (refer to Anaphylaxis section). The main underlying mechanism is type I hypersensitivity reactions by interaction of allergens within the donor plasma with preformed immunoglobulin (Ig) E antibodies in the recipient (62,64).

Prompt cessation of transfusion and administration of antihistamine agent are required for symptomatic relief (21,65). Premedication with antihistamine agents for reducing the incidence of ATRs has been shown to have suboptimal efficacy (66,67). For patients with a history of ATRs, close monitoring during subsequent transfusions is required (12).

Two observational studies in the US showed that PAS-stored apheresis platelets could reduce the incidence of allergic reactions by about 50%: ATR incidences of 1.01% for PAS-platelets versus 1.85% for platelets in plasma from the study by Tobian *et al.* (26), and incidences of 0.29% for platelets in PAS-C versus 0.82% for platelets in plasma from the study by Cohn *et al.* (62).

Platelet washing has also been shown to be beneficial for preventing ATRs (50,51,68-70). One observational study in Japan revealed that ATR to apheresis platelets decreased after the release of washed PCs: ATR incidences of 0.36% for washed PCs versus 3.44% for unwashed PCs (51).

### FNHTR

FNHTRs, clinically presented as a temperature increase (above 38 °C or more than 1 °C increase above baseline) during or shortly after transfusion, are also frequent adverse events, which occur in 4% to 6% of platelet transfusions (23,62). Transient symptoms such as chills, rigors, tachypnea, anxiety, and headache can be accompanied, which are not specific to FNHTRs, hence FNHTRs should be diagnosed by exclusion (21). In case of developing fever, immediate cessation of platelet transfusion and assessment of infection or hemolysis are required (12,71) (refer to TTBI section). Fever can be managed with antipyretics, but premedication to prevent FNHTRs has been reported to be ineffective (72-75).

Multiple factors such as donor leucocytes, cytokine accumulation during storage and leucoagglutinins in the donor or recipient are thought to cause FNHTRs (23,64,76). One observational study in the US showed that prestorage leukoreduction of buffy-coat pooled whole-blood-derived

platelets is effective enough to decrease FNHTR risk by about 90%: FNHTR incidence of 0.15% for leukoreduced pooled platelets versus 2.18% for untreated pooled platelets (77). Additionally, both PAS-stored apheresis platelets and whole-blood-derived platelets have been shown to reduce FNHTR rate compared to plasma platelets (62,78). Washed platelets have also been reported to efficiently mitigate FNHTRs (79), but in an observational study in Japan, washing of single-donor apheresis platelets had no effect on reducing FNHTR incidence (51). Hence, the exact effect of platelet washing on reducing FNHTRs is controversial (63).

### Management of severe adverse events related to platelet transfusions

Severe adverse events associated with platelet transfusions include TTBI, anaphylaxis, respiratory distress including transfusion-related acute lung injury (TRALI) and transfusion-associated circulatory overload (TACO) (23,80,81).

### TTBI

Bacterial infection has long been the highest TTI risk associated with platelet transfusion (23,47,82). Platelets are stored at room temperature mostly due to negative impact of cold storage on platelet function and survival (83-85), which poses platelet components at a high risk of bacterial contamination compared to RBC and plasma component (31,36,86,87). Septic transfusion reactions caused by TTBI are of great concern due to the fatality and severity even if transfusion-related infectious complications are less common than noninfectious complications (44,54,55).

Since the bacterial infection risk increases in relationship to the storage duration, the shelf-life of platelets stored at room temperature is generally shorter than that of other components, with most countries adopting the 5 days. Recently, the large-volume, delayed-sampling bacterial screening algorithm has been implemented in the UK and in Canada (32,88), which significantly reduced the risk of bacterial transmission, and allowed the extension of the PC shelf-life to 7 days. The US Food and Drug Administration also approved the 7-day extension of the PC shelf-life, provided a secondary bacterial testing, through the conventional testing or by a rapid test, is performed, or the large-volume, delayed-sampling bacterial screening algorithm is applied (89). Estimates of TTBI incidence with



platelet transfusions are variable between pooled platelets and single-donor apheresis platelets, ranging from 10 to 70 in million units (87,90), and the incidence is estimated to be higher than incidence of transfusion-transmitted human immunodeficiency virus (HIV) or hepatitis C virus (HCV) infections (about one in two million units transfused) (91,92).

Typical clinical manifestations of TTBI include fever, rigors, tachycardia, hypertension and hypotension (23,87), which require differentiation from ATRs, FNHTRs and other causes. In some cases, abdominal pain, back pain, nausea, vomiting and hypothermia can be presented. The duration between completion of transfusion and appearance of related symptoms is short with a median of 30 minutes, depending on multiple factors such as bacterial inoculum size, bacterial virulence and immune status of the recipient (87,90).

Once TTBI is suspected, prompt cessation of transfusion and evaluation for the presence of bacterial contamination in all transfused products are required (93). Bacterial cultures should be taken before empiric treatment with broad-spectrum antibiotics is initiated (65). Antibiotic treatments should be adjusted based on subsequent culture results.

Due to the low sensitivity of surrogate markers such as pH, glucose and/or platelet swirling for bacterial detection, post-collection automated culture methods have been widely implemented in developed countries (23,94). Bacterial screening by automated culture systems such as BacT/Alert and eBDS has shown increased sensitivity for bacterial detection (71,95-101). Another option for mitigating bacterial contamination is rapid immunoassay, which can detect bacterial wall components. Combination of bacterial culture and immunoassay tests has been shown to increase sensitivity for bacterial detection, however, there still remains a residual TTBI risk (71,82,102,103).

PRT, a method relying on an ultraviolet photo activation system, is an available option for pathogen reduction in platelets. It is currently used to varying degrees in most developed countries (*Table 1*) (9,10,23), which have been shown to contribute to decrease of fatality by TTBI (46). Although recent reports have shown that current PRT can decrease platelet recovery following transfusion (34,104), PRT might also be beneficial for risk reduction of agents where screening tests do not exist or have poor sensitivity (10,104,105).

In Japan, where single-donor apheresis platelets are transfused under “3-day limitation policy” with no bacterial

culture testing, investigation of 86 suspected TTBI cases during 2007 to 2018 revealed that the short shelf-life of PC is associated with a low TTBI incidence: one, six and ten TTBI cases were caused by Day 1, 2 and 3 PCs, respectively (36). From the hemovigilance data provided in *Table 5*, incidence of suspected and confirmed bacterial contamination is as low as 0.46 and 0.03 per 100,000 transfusion bags (including PC, RBC, and FFP). Under the circumstance, visual inspection of PC bags prior to and during the transfusion for identifying aggregates in the bag or filter clogging seems to be crucial in preventing TTBI.

### *Anaphylaxis*

Anaphylactic reactions, the most severe ATRs occurring in eight per 100,000 units, usually present as respiratory distress, bronchospasm and hypotension (106). Immediate cessation of transfusion and prompt intramuscular epinephrine administration with supportive care are required for treatment of anaphylactic reactions (65,72,107). Other medications to be considered are H<sub>1</sub> antihistamine agents, bronchodilators, glucocorticoid and H<sub>2</sub> antihistamine agents (65). For patients with past history of anaphylactic reactions, specific serum protein deficiency such as IgA and haptoglobin should be investigated (65).

For IgA-deficient patients who have anti-IgA antibodies without previous anaphylactic reactions, transfusion of IgA-deficient or washed blood components can be applied (65,108). In Japan, after the release of washed PCs by the JRCS in 2016, no severe allergic reactions to washed single-donor-apheresis PCs have been reported (51).

### *TRALI*

TRALI is a rare but potentially fatal respiratory complication of blood component transfusion, which is caused by activated pulmonary neutrophils (12,109-111). Traditionally, TRALI was defined as newly-developed pulmonary distress within 6 hours of completion of transfusion when the recipient had no other risk factors for acute respiratory distress syndrome (111). Recipient risk factors for TRALI are liver transplantation surgery, chronic alcohol abuse, shock, higher peak airway pressure while being mechanically ventilated, current smoking, higher interleukin (IL)-8 levels and positive fluid balance, whereas blood component risk factors for TRALI are high-plasma-volume blood components, increased volume of transfused HLA class II antibody or anti-human neutrophil

antigen (HNA) antibody (23,112-114). Although TRALI had been the leading cause of transfusion-associated fatalities, occurring in 1 in 3,891 units transfused (115), implementation of mandated strategies such as deferral of multiparous female donors was successful in the risk reduction with the incidence of 1 in 12,345 units (114).

Development of immune TRALI had been thought to fit for a “two-hit” model, in which the first hit is a clinical disorder that causes activation of the pulmonary endothelium, leading to the sequestration and priming of neutrophils in the lung, and the second hit is the transfusion of blood components, especially plasma (116). However, as TRALI can also occur in the absence of demonstrable antibodies, recently-proposed “multicausal model” might better describe TRALI pathogenesis (117).

Despite recently revised definition of TRALI (118), accurate diagnosis of TRALI is still challenging. Clinical symptoms of TRALI include dyspnea, tachypnea, and hypoxemia, sometimes accompanied by rigors, tachycardia, fever, hypothermia, and hypotension or hypertension, which require differentiation from TACO, anaphylaxis and septic reaction (110,114). Treatment of TRALI is mainly supportive care with supplemental oxygen or mechanical ventilation as needed. Additionally, both fluid and transfusion restriction can be preventive (12,114).

In Japan, since 2004, suspected cases of TRALI are submitted to the JRCS, where a case record auditing by lung specialist physicians is conducted to evaluate if the clinical and radiological findings meet the criteria of TRALI. In addition, for the further investigation of the adverse event, various types of tests, including HLA and HNA antibody screening, peripheral blood leukocyte crossmatch, measurement of N-terminal prohormone of brain natriuretic peptide (NT-proBNP), and screening for anti-plasma proteins and plasma protein deficiencies, are conducted (39,119,120). The annual numbers of confirmed TRALI and p-TRALI cases from 2010 to 2018 were 4–14 and 1–15, respectively, and in the recent years, a decreasing trend is observed (*Table 5*). Among the 120 cases of TRALI and p-TRALI diagnosed during the period, four cases were fatal (121).

## TACO

TACO is the most frequent but underdiagnosed and underreported pulmonary complication of any blood component transfusion, including platelets (23,110,122). It

typically occurs in patients receiving transfusion of a large volume of blood over a short period of time, or in those with underlying diseases (12,123,124). Risk factors for TACO have been reported to include cardiac, pulmonary, or renal disease, older age (over 70 years), low body weight, and pre-transfusion positive fluid balance (123-126), however, other factors including inflammation are supposed to cause TACO (110). The estimated frequency of TACO varies from 1% to 11%, depending on clinical settings (110,127-129). The recent reports from UK and US showed that TACO is one of the leading causes of transfusion-related fatalities (54,55).

The definition of TACO has recently been revised by the ISBT, the International Hemovigilance Network, and AABB, requiring new onset or acute exacerbation of three or more of the following, within 12 hours of transfusion: respiratory distress, raised brain natriuretic peptide (BNP or NT-pro-BNP), increased central venous pressure, left heart failure, positive fluid balance, or pulmonary edema (12,126). Despite the revised diagnostic criteria, precise diagnosis of TACO is mostly difficult due to overlapping of clinical symptoms with TRALI, septic transfusion reaction and acute HTR (12).

Treatment of TACO requires prompt cessation of transfusion, oxygen supplementation, and administration of diuretics as needed (125,130). For at-risk patients, slow infusion rates and pre-transfusion administration of diuretics can reduce the incidence of TACO (12,23,130). The benefit of diuretics before or during the transfusion for patients with a history of TACO is being investigated (131,132).

Since 2012, when the JRCS started evaluating the suspected TACO cases, the number of definite TACO cases has increased year by year, reaching 55 cases in 2018 (*Table 5*). Among the 798 suspected cases of TRALI, 309 were classified as TACO, but the remaining 369 cases were classified as cardiogenic pulmonary edema (*Table 5*), which suggests that circulatory overload may be overlooked in the clinical practice (39).

## Other adverse events to be considered

Other adverse events to be considered are HTR, TTI other than TTBI, transfusion-associated graft versus host disease (TA-GVHD) and post-transfusion purpura (PTP). Platelet transfusion refractoriness due to alloimmunization is also of concern, which is reviewed in another article in the same

issue of *Annals of Blood* journal.

### HTR

As with the cases with TRALI, TACO, FNHTRs and ATRs, amounts of incompatible donor plasma in PC bags can cause hemolysis in recipients (23). Clinical manifestation of acute HTRs associated platelet transfusions is less severe than those with incompatible RBCs (133). Since platelets are often transfused without respecting the ABO compatibility in most Western countries due to the limited stock availability (81), a sizable number of minor ABO-incompatible platelet products are routinely issued (133-136). Although minor ABO-incompatible platelet transfusions rarely cause recipient intravascular hemolysis, underreporting of HTRs due to minor ABO-incompatible platelet transfusions is of concern (135,137). Screening of platelet components for titers of anti-A and anti-B antibodies is a mitigation strategy for recipient hemolysis, but consensus on methods or thresholds for “high-titer” is not well established so far (23,138).

Even if ABO-identical platelet transfusions result in a higher platelet increment, the role of ABO matching in platelet transfusions is still an issue of debate (139,140). Considering a high burden of issuing ABO-identical platelets (140), other mitigation strategies such as utilization of PAS-PC or washed PC seem to be more reasonable to minimize the risk of ABO-minor incompatible reactions.

In the Japanese practice, ABO-incompatible PCs can be supplied only in case there is a need to prioritize HLA-matching. But even in such cases, blood group O PC is never supplied to patients of other blood group types, and antibody titers are measured and only those with low antibody titers are supplied in case of major mismatch between donor and recipient, e.g., blood group A PC to a blood group B patient. Recent hemovigilance data of Japan showed that all recent cases of HTRs associated platelet transfusions were due to the administration of ABO-incompatible HLA-matched platelets, with an incidence of 0.0021% (1 per 47,800 HLA-matched PC bags) (42).

### TTI other than TTBI

Improvement of donor testing over the past few decades, including nucleic acid-amplification technologies (NATs), has contributed to significant reduction of TTIs by HIV, hepatitis B virus (HBV) and HCV (23,28). Current

estimates of these TTI rates are reported to be 1 in 2 to 3 million units (28).

Adoption of NAT screening for other viruses such as West Nile virus, Zika virus, and HEV varies depending on the country or the region (141-149). Leukoreduction has been shown to reduce risks of transfusion-related transmission of cytomegalovirus, human T-lymphotropic virus and human herpesvirus 8 (23,150-152).

In Japan, NAT testing of HIV, HBV and HCV was implemented in 1999, initially with a mini-pool of 500, then reduced to 50 in 2000, to 20 in 2004, and 10 years later, in 2014, the individual NAT was implemented. Recent estimated risk of HBV transmission is 1 in 2 million units, and those of HCV and HIV transmission are immeasurably small (35). Selective screening of *Trypanosoma cruzi* and individual NAT testing for HEV started in August 2016 and in August 2020, respectively. Testing for *Babesia microti*, West Nile virus and Zika virus are not routinely performed.

### TA-GVHD

TA-GVHD is an extremely rare adverse event caused by transfusion of cellular components containing viable donor lymphocytes that engraft in the transfused recipient and attack the recipient's organs (12,23,153). The mortality rate of TA-GVHD is as high as 90%, and blood products from biologically related donors (partial HLA matching) and immunocompromised recipients are two major risk factors for TA-GVHD (23,154,155).

Within 5 to 10 days after transfusion, erythema, fever, abdominal pain, diarrhea, nausea, and vomiting develop with abnormal laboratory results such as pancytopenia, abnormal liver function, and electrolyte disturbances. Subsequently, bone marrow aplasia develops within 21 days. Most of fatal TA-GVHD cases are attributable to infections (154).

The preferred prevention of TA-GVHD is irradiation of cellular blood components with gamma-rays or X-rays, which has been successful in dramatic decrease of TA-GVHD incidence (153,155-157). PRTs have also been considered as an alternative method to prevent TA-GVHD development in some countries (154,158).

As the highest incidence of TA-GVHD had been reported in Japan, the supply of irradiated blood products was introduced nationwide by the JRCS in 1998, reflecting the revised guidelines for the irradiation of blood to prevent post-transfusion GVHD by the Japan Society of Blood

Transfusion (former JSTMCT), which recommended transfusion of irradiated blood for at-risk patients (157). Thereafter, in principle, irradiation of all cellular components is recommended in the present guidelines for the appropriate use of blood products released by the Japanese Ministry of Health, Labor and Welfare. Hospitals may request irradiated blood to the JRCS or irradiate at their own service prior to transfusion. No TA-GVHD cases by blood components distributed by the JRCS have been confirmed in Japan since 2000 (40,156).

### PTP

PTP is a rare bleeding disorder caused by alloantibodies specific to platelet antigens. It mostly affects HPA-1a-negative individuals who have been alloimmunized by previous pregnancy and/or transfusion while other HPA antigens might be involved (12,159). Typically, thrombocytopenia is severe but resolves spontaneously within several weeks. The diagnosis is confirmed by the detection of platelet specific alloantibodies. PTP is treated with intravenous Ig, corticosteroids, and plasmapheresis (12,159). To prevent PTP recurrence, HPA-compatible platelets are indicated (160).

### Limitations and future perspective

This article has several limitations. First, the incidence of each type of adverse reaction, referred from the literatures outside Japan, might be affected by the inter-country difference of reporting systems. Second, the difference of reporting systems and participants might limit the direct comparison of the two hemovigilance data of Japan. And underreporting might affect both collected data due to their nature of passive surveillance. Considering each advantage for collecting transfusion-related adverse events, integration of two hemovigilance systems seems reasonable. Third, the descriptions related to transfusion practice in Japan could be insufficient, as the number of literatures from Japan is limited. Fourth, this article could not cover all literatures outside Japan regarding adverse effects to platelet transfusion due to the nature of narrative review.

Implementation of bacterial culture screening and PRT, and supply of PAS-PC are under consideration in Japan. For ensuring safe and appropriate blood transfusion in Japan, both observational and investigational researches should be encouraged.

### Summary

The unique situation of transfusing leukoreduced single-donor apheresis platelets in Japan is highlighted as the short shelf-life of 3 days and visual inspection before issue, with no implementation of bacterial culture screening and pathogen reduction technologies. From two recent hemovigilance data of Japan during 2010 to 2018, non-HTRs including ATRs and FNHTRs are the most frequent adverse events related to platelet transfusions in Japan, with an incidence of 3.16%. The incidence is also higher than that with RBC (0.64%) and plasma transfusions (1.11%). Severe adverse events related to any of blood components are anaphylaxis, respiratory distress including TRALI and TACO, and TTIs, with the incidence of, 7.31, 1.38, and 0.17 per 100,000 transfusion bags, respectively. During the period, a total of six fatal cases were reported, which included one case of TTBI by *E. coli*, one case of HEV infection, 3 cases of TRALI, and one case of p-TRALI. Even under the unique situations in Japan, incidence of adverse events related to platelet transfusions is mostly consistent with previous related reports outside Japan.

Various risk mitigation strategies have been successful for securing safety of platelet transfusions, however, there is still room for further improvement. This can be achieved by patient blood management strategies, including appropriate use of blood products, with adherence to evidence-based transfusion guidelines, minimizing unnecessary transfusions and regularly performing reflection on transfusion practices. ATRs including anaphylaxis can be mostly mitigated by the use of PAS reposition or washed platelets. Bacterial screening of PC and/or PRT are being applied for the prevention of TTBI, and good results have been shown with the large-volume, delayed-sampling bacterial screening algorithm implemented in the UK and Canada, which may be superior to the strategy applied in Japan. TRALI and TACO are conditions that can be prevented or treated, provided the condition is appropriately recognized and managed. Thus, it is essential that clinicians recognize the adverse events that may occur during or after the platelet transfusion, and understand the appropriate management of each condition.

### Acknowledgments

The authors thank Dr Isao Hamaguchi of the National Institute of Infectious Diseases for providing hemovigilance data (53), and Prof Hitoshi Okazaki of the University of



Tokyo hospital for helpful comment.

*Funding:* None.

## Footnote

*Provenance and Peer Review:* This article was commissioned by the Guest Editor (Pilar Solves) for the series “Platelet Transfusion” published in *Annals of Blood*. The article has undergone external peer review.

*Reporting Checklist:* The authors have completed the Narrative Review reporting checklist. Available at <http://dx.doi.org/10.21037/aob-20-90>

*Peer Review File:* Available at <http://dx.doi.org/10.21037/aob-20-90>

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/aob-20-90>). The series “Platelet Transfusion” was commissioned by the editorial office without any funding or sponsorship. Dr. NHT serves as an unpaid editorial board member of *Annals of Blood*. The authors have no other conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

## References

- Kaufman RM, Djulbegovic B, Gernsheimer T, et al. Platelet transfusion: A clinical practice guideline from the AABB. *Ann Intern Med* 2015;162:205-13.
- Blumberg N, Heal JM, Phillips GL. Platelet transfusions: Trigger, dose, benefits, and risks. *F1000 Med Rep* 2010;2:5.
- Connell NT. Transfusion Medicine. *Prim Care* 2016;43:651-9.
- Greeno E, McCullough J, Weisdorf D. Platelet utilization and the transfusion trigger: A prospective analysis. *Transfusion* 2007;47:201-5.
- Estcourt LJ. Why has demand for platelet components increased? A review. *Transfus Med* 2014;24:260-8.
- Charlton A, Wallis J, Robertson J, et al. Where did platelets go in 2012? A survey of platelet transfusion practice in the North of England. *Transfus Med* 2014;24:213-8.
- Cameron B, Rock G, Olberg B, et al. Evaluation of platelet transfusion triggers in a tertiary-care hospital. *Transfusion* 2007;47:206-11.
- Storch EK, Custer BS, Jacobs MR, et al. Review of current transfusion therapy and blood banking practices. *Blood Rev* 2019;38:100593.
- Capocelli KE, Dumont LJ. Novel platelet storage conditions : additive solutions, gas, and cold. *Curr Opin Hematol* 2014;21:491-6.
- Waters L, Cameron M, Padula MP, et al. Refrigeration, cryopreservation and pathogen inactivation: an updated perspective on platelet storage conditions. *Vox Sang* 2018;113:317-28.
- Bolton-Maggs PH. Bullet points from SHOT: key messages and recommendations from the Annual SHOT Report 2013. *Transfus Med* 2014;24:197-203.
- Delaney M, Wendel S, Bercovitz RS, et al. Transfusion reactions: prevention, diagnosis, and treatment. *Lancet* 2016;388:2825-36.
- Domen RE, Hoeltge GA. Allergic transfusion reactions: An evaluation of 273 consecutive reactions. *Arch Pathol Lab Med* 2003;127:316-20.
- Bishop D, Tandy N, Anderson N, et al. A clinical and laboratory study of platelet concentrates produced by pooled buffy coat and single donor apheresis technologies. *Transfus Sci* 1995;16:187-8.
- Tormey CA, Sweeney JD, Champion MH, et al. Analysis of transfusion reactions associated with prestorage-pooled platelet components. *Transfusion* 2009;49:1242-7.
- Enright H, Davis K, Gernsheimer T, et al. Factors influencing moderate to severe reactions to PLT transfusions: Experience of the TRAP multicenter clinical trial. *Transfusion* 2003;43:1545-52.
- ISBT Working Party on Haemovigilance. Proposed standard definitions for surveillance of non-infectious adverse transfusion reactions [cited 2020 Dec 3]. 2011. Available online: <https://www.isbtweb.org/fileadmin/>

- user\_upload/Proposed\_definitions\_2011\_surveillance\_non\_infectious\_adverse\_reactions\_haemovigilance\_incl\_TRALI\_correction\_2013.pdf
18. Savage WJ, Tobian AA, Fuller AK, et al. Allergic transfusion reactions to platelets are associated more with recipient and donor factors than with product attributes. *Transfusion* 2011;51:1716-22.
  19. Heddle NM, Klama LN, Griffith L, et al. A prospective study to identify the risk factors associated with acute reactions to platelet and red cell transfusions. *Transfusion* 1993;33:794-7.
  20. Tobian AAR, Savage WJ, Tisch DJ, et al. Prevention of allergic transfusion reactions to platelets and red blood cells through plasma reduction. *Transfusion* 2011;51:1676-83.
  21. Goel R, Tobian AAR, Shaz BH. Noninfectious transfusion-associated adverse events and their mitigation strategies. *Blood* 2019;133:1831-9.
  22. Pietersz RNI, Reesink HW, Panzer S, et al. Bacterial contamination in platelet concentrates. *Vox Sang* 2014;106:256-83.
  23. Katus MC, Szczepiorkowski ZM, Dumont LJ, et al. Safety of platelet transfusion: Past, present and future. *Vox Sang* 2014;107:103-13.
  24. van der Meer PF, de Korte D. Platelet Additive Solutions: A Review of the Latest Developments and Their Clinical Implications. *Transfus Med Hemother* 2018;45:98-102.
  25. Kacker S, Ness PM, Savage WJ, et al. The cost-effectiveness of platelet additive solution to prevent allergic transfusion reactions. *Transfusion* 2013;53:2609-18.
  26. Tobian AAR, Fuller AK, Uglik K, et al. The impact of platelet additive solution apheresis platelets on allergic transfusion reactions and corrected count increment (CME). *Transfusion* 2014;54:1523-9.
  27. Alhumaidan H, Sweeney J. Current Status of Additive Solutions for Platelets. *J Clin Apher* 2012;27:93-8.
  28. Busch MP, Bloch EM, Kleinman S. Prevention of transfusion-transmitted infections. *Blood* 2019;133:1854-64.
  29. Thyer J, Perkowska-Guse Z, Ismay SL, et al. Bacterial testing of platelets - has it prevented transfusion-transmitted bacterial infections in Australia? *Vox Sang* 2018;113:13-20.
  30. Levy JH, Neal MD, Herman JH. Bacterial contamination of platelets for transfusion: strategies for prevention. *Crit Care* 2018;22:271.
  31. Wagner SJ. Transfusion-transmitted bacterial infection: Risks, sources and interventions. *Vox Sang* 2004;86:157-63.
  32. McDonald C, Allen J, Brailsford S, et al. Bacterial screening of platelet components by National Health Service Blood and Transplant, an effective risk reduction measure. *Transfusion* 2017;57:1122-31.
  33. Stubbs JR, Homer MJ, Silverman T, et al. The current state of the platelet supply in the US and proposed options to decrease the risk of critical shortages. *Transfusion* 2021;61:303-12.
  34. Estcourt LJ, Malouf R, Hopewell S, et al. Pathogen-reduced platelets for the prevention of bleeding. *Cochrane Database Syst Rev* 2017;7:CD009072.
  35. The Japan Red Cross Society. Transfusion Information 1804-159 [Internet]. 2018 [cited 2020 Dec 13]. Available online: [http://www.jrc.or.jp/mr/english/pdf/yuketsu\\_johou\\_1804\\_159.pdf](http://www.jrc.or.jp/mr/english/pdf/yuketsu_johou_1804_159.pdf)
  36. Satake M, Kozakai M, Matsumoto M, et al. Platelet safety strategies in Japan: impact of short shelf life on the incidence of septic reactions. *Transfusion* 2020;60:731-8.
  37. Takami A, Matsushita T, Ogata M, et al. Guideline for the use of platelet transfusion concentrates based on scientific evidence: update 2019. *Japanese Journal of Transfusion and Cell Therapy* 2019;65:544-61.
  38. Odaka C, Kato H, Otsubo H, et al. Online reporting system for transfusion-related adverse events to enhance recipient haemovigilance in Japan: A pilot study. *Transfus Apher Sci* 2013;48:95-102.
  39. Satake M. Experience of haemovigilance in Japan. *ISBT Sci Ser* 2018;13:298-301.
  40. The Japan Red Cross Society. Transfusion information [Internet]. [cited 2020 Dec 3]. Available online: <http://www.jrc.or.jp/mr/news/transfusion/>
  41. Omokawa S, Abe M. The plasma supply in Japan. *Transfus Apher Sci* 2020;59:102749.
  42. Sato T, Goto N, Tasaki T. Hemolytic Transfusion Reactions. *N Engl J Med* 2019;381:1396-7.
  43. Blood Products Advisory Committee November 30 - December 1, 2017 Meeting Final Agenda [Internet]. 2017 [cited 2020 Dec 3]. Available online: <https://www.fda.gov/media/111165/download>
  44. Canadian Blood Services. Surveillance Report [Internet]. 2017 [cited 2020 Dec 3]. Available online: [https://professionaleducation.blood.ca/sites/msi/files/surveillance\\_report\\_2017\\_final.pdf](https://professionaleducation.blood.ca/sites/msi/files/surveillance_report_2017_final.pdf)
  45. NZBS. Haemovigilance annual report, 2015-19 [Internet]. 2019 [cited 2020 Dec 3]. Available online: <https://www.nzblood.co.nz/clinical-information/haemovigilance-programme/haemovigilance-annual-report-2012/>
  46. Benjamin RJ, Kline L, Dy BA, et al. Bacterial



- contamination of whole blood-derived platelets: The introduction of sample diversion and prestorage pooling with culture testing in the American Red Cross. *Transfusion* 2008;48:2348-55.
47. Ramirez-Arcos S, DiFranco C, McIntyre T, et al. Residual risk of bacterial contamination of platelets: six years of experience with sterility testing. *Transfusion* 2017;57:2174-81.
  48. Savage WJ, Savage JH, Tobian AAR, et al. Allergic agonists in apheresis platelet products are associated with allergic transfusion reactions. *Transfusion* 2012;52:575-81.
  49. Shimada E, Tadokoro K, Watanabe Y, et al. Anaphylactic transfusion reactions in haptoglobin-deficient patients with IgE and IgG haptoglobin antibodies. *Transfusion* 2002;42:766-73.
  50. Hirayama F. Current understanding of allergic transfusion reactions: Incidence, pathogenesis, laboratory tests, prevention and treatment. *Br J Haematol* 2013;160:434-44.
  51. Ikebe E, Matsuoka S, Tanaka A, et al. Reduction in adverse transfusion reactions with increased use of washed platelet concentrates in Japan—A retrospective multicenter study. *Transfus Apher Sci* 2019;58:162-8.
  52. Okazaki H. The benefits of the Japanese haemovigilance system for better patient care. *ISBT Sci Ser* 2007;2:104-9.
  53. The Japan Society of Transfusion Medicine and Cell Therapy. Hemovigilance data of adverse transfusion reactions [Internet]. [cited 2020 Dec 3]. Available online: [http://yuketsu.jstmct.or.jp/medical/side\\_effect/](http://yuketsu.jstmct.or.jp/medical/side_effect/)
  54. Fatalities Reported to FDA Following Blood Collection and Transfusion: Annual Summary for FY2018 [Internet]. 2018 [cited 2020 Dec 7]. Available online: <https://www.fda.gov/vaccines-blood-biologics/report-problem-center-biologics-evaluation-research/transfusiondonation-fatalities>
  55. Serious Hazards of Transfusion (SHOT). Serious Hazards of Transfusion annual reports and summaries 1996-2017 [Internet]. [cited 2020 Dec 3]. Available online: <https://www.shotuk.org/shot-reports/>
  56. Muylle L, Roisin T. Guidelines of the Belgian Hemovigilance Report. *Belgian J Hematol* 2010;1:57-61.
  57. Rieux C, Brittenham G, Bachir D, et al. Delayed hemolytic transfusion reaction in the French hemovigilance system. *Transfus Clin Biol* 2019;26:109-11.
  58. Jutzi M, Mansouri Taleghani B, Rueesch M, et al. Nationwide Implementation of Pathogen Inactivation for All Platelet Concentrates in Switzerland. *Transfus Med Hemother* 2018;45:151-6.
  59. National Blood Authority Australia. Australian Haemovigilance Report, 2013-14 to 2017-18 [Internet]. [cited 2020 Dec 4]. Available online: <https://www.blood.gov.au/haemovigilance-reporting>
  60. Hong H, Xiao W, Lazarus HM, et al. Detection of septic transfusion reactions to platelet transfusions by active and passive surveillance. *Blood* 2016;127:496-502.
  61. Cohn CS, Delaney M, Johnson ST, et al. Technical Manual. 20th edition. Bethesda (MD): AABB, 2020.
  62. Cohn CS, Stubbs J, Schwartz J, et al. A comparison of adverse reaction rates for PAS C versus plasma platelet units. *Transfusion* 2014;54:1927-34.
  63. Harvey AR, Basavaraju SV, Chung KW, et al. Transfusion-related adverse reactions reported to the National Healthcare Safety Network Hemovigilance Module, United States, 2010 to 2012. *Transfusion* 2015;55:709-18.
  64. Garraud O, Tariket S, Sut C, et al. Transfusion as an inflammation Hit: Knowns and Unknowns. *Front Immunol* 2016;7:534.
  65. Tinegate H, Birchall J, Gray A, et al. Guideline on the investigation and management of acute transfusion reactions Prepared by the BCSH Blood Transfusion Task Force. *Br J Haematol* 2012;159:143-53.
  66. Geiger TL, Howard SC. Acetaminophen and Diphenhydramine Premedication for Allergic and Febrile Nonhemolytic Transfusion Reactions: Good Prophylaxis or Bad Practice? *Transfus Med Rev* 2007;21:1-12.
  67. Ning S, Solh Z, Arnold DM, et al. Premedication for the prevention of nonhemolytic transfusion reactions: a systematic review and meta-analysis. *Transfusion* 2019;59:3609-16.
  68. Cholette JM, Henrichs KF, Alfieri GM, et al. Washing red blood cells and platelets transfused in cardiac surgery reduces postoperative inflammation and number of transfusions: Results of a prospective, randomized, controlled clinical trial. *Pediatr Crit Care Med* 2012;13:290-9.
  69. Oikawa S, Minegishi M, Endo K, et al. Washing platelets twice with a bicarbonated Ringer's solution significantly reduces plasma protein levels while maintaining platelet quality. *Transfus Apher Sci* 2016;55:344-6.
  70. Fujiwara SI, Fujishima N, Kanamori H, et al. Released washed platelet concentrates are effective and safe in patients with a history of transfusion reactions. *Transfus Apher Sci* 2018;57:746-51.
  71. Jacobs MR, Smith D, Heaton WA, et al. Detection of bacterial contamination in prestorage culture-negative apheresis platelets on day of issue with the Pan Genera

- Detection test. *Transfusion* 2011;51:2573-82.
72. Martí-Carvajal AJ, Solà I, González LE, et al. Pharmacological interventions for the prevention of allergic and febrile non-haemolytic transfusion reactions. *Cochrane Database Syst Rev* 2010;2010:CD007539.
  73. Wang SE, Lara PN, Lee-Ow A, et al. Acetaminophen and diphenhydramine as premedication for platelet transfusions: A prospective randomized double-blind placebo-controlled trial. *Am J Hematol* 2002;70:191-4.
  74. Kennedy LD, Case LD, Hurd DD, et al. A prospective, randomized, double-blind controlled trial of acetaminophen and diphenhydramine pretransfusion medication versus placebo for the prevention of transfusion reactions. *Transfusion* 2008;48:2285-91.
  75. Sanders RP, Maddirala SD, Geiger TL, et al. Premedication with acetaminophen or diphenhydramine for transfusion with leucoreduced blood products in children. *Br J Haematol* 2005;130:781-7.
  76. Vamvakas EC. Relative safety of pooled whole blood-derived versus single-donor (apheresis) platelets in the United States: A systematic review of disparate risks. *Transfusion* 2009;49:2743-58.
  77. Paglino JC, Pomper GJ, Fisch GS, et al. Reduction of febrile but not allergic reactions to RBCs and platelets after conversion to universal prestorage leukoreduction. *Transfusion* 2004;44:16-24.
  78. Heddle NM, Blajchman MA, Meyer RM, et al. A randomized controlled trial comparing the frequency of acute reactions to plasma-removed platelets and prestorage WBC-reduced platelets. *Transfusion* 2002;42:556-66.
  79. Vo TD, Cowles J, Heal JM, et al. Platelet washing to prevent recurrent febrile reactions to leucocyte-reduced transfusions. *Transfus Med* 2001;11:45-7.
  80. Garraud O, Cognasse F. Could platelet washing be used to reduce adverse reactions in patients receiving platelet component transfusions? *Ann Blood* 2019;4:9.
  81. Solves Alcaina P. Platelet Transfusion: And Update on Challenges and Outcomes. *J Blood Med* 2020;11:19-26.
  82. Rios J, Westra J, Dy B, et al. Adoption trends of point of issue Verax PGD rapid test for bacterial screening of platelets between 2013 and 2018 among hospitals supplied by the American Red Cross and impact on platelet availability. *Transfusion* 2020;60:1364-72.
  83. Rumjantseva V, Grewal PK, Wandall HH, et al. Dual roles for hepatic lectin receptors in the clearance of chilled platelets. *Nat Med* 2009;15:1273-80.
  84. Hoffmeister KM, Josefsson EC, Isaac NA, et al. Glycosylation restores survival of chilled blood platelets. *Science* 2003;301:1531-4.
  85. Hoffmeister KM, Felbinger TW, Falet H, et al. The clearance mechanism of chilled blood platelets. *Cell* 2003;112:87-97.
  86. Brecher ME, Hay SN. Bacterial contamination of blood components. *Clin Microbiol Rev* 2005;18:195-204.
  87. Kuehnert MJ, Roth VR, Haley NR, et al. Transfusion-transmitted bacterial infection in the United States, 1998 through 2000. *Transfusion* 2001;41:1493-9.
  88. Ramirez-Arcos S, Evans S, McIntyre T, et al. Extension of platelet shelf life with an improved bacterial testing algorithm. *Transfusion* 2020;60:2918-28.
  89. Bacterial Risk Control Strategies for Blood Collection Establishments and Transfusion Services to Enhance the Safety and Availability of Platelets for Transfusion | FDA [Internet]. [cited 2020 Dec 10]. Available online: <https://www.fda.gov/regulatory-information/search-fda-guidance-documents/bacterial-risk-control-strategies-blood-collection-establishments-and-transfusion-services-enhance>
  90. Perez P, Rachid Salmi L, Folléa G, et al. Determinants of transfusion-associated bacterial contamination: Results of the French BACTHEM case-control study. *Transfusion* 2001;41:862-72.
  91. Goodnough LT, Shander A, Brecher ME. Transfusion medicine: Looking to the future. *Lancet* 2003;361:161-9.
  92. Vasconcelos E, Seghatchian J. Bacterial contamination in blood components and preventative strategies: An overview. *Transfus Apher Sci* 2004;31:155-63.
  93. Eder AF, Goldman M. How do I investigate septic transfusion reactions and blood donors with culture-positive platelet donations? *Transfusion* 2011;51:1662-8.
  94. Yomtovian R, Brecher ME. pH and glucose testing of single-donor apheresis platelets should be discontinued in favor of a more sensitive detection method. *Transfusion* 2005;45:646-8.
  95. Dumont LJ, Kleinman S, Murphy JR, et al. Screening of single-donor apheresis platelets for bacterial contamination: The PASSPORT study results. *Transfusion* 2010;50:589-99.
  96. Pearce S, Rowe GP, Field SP. Screening of platelets for bacterial contamination at the Welsh Blood Service. *Transfus Med* 2011;21:25-32.
  97. Larsen CP, Ezligini F, Hermansen NO, et al. Six years' experience of using the BacT/ALERT system to screen all platelet concentrates, and additional testing of outdated platelet concentrates to estimate the frequency of false-negative results. *Vox Sang* 2005;88:93-7.

98. Walther-Wenke G, Däubener W, Heiden M, et al. Effect of safety measures on bacterial contamination rates of blood components in Germany. *Transfus Med Hemother* 2011;38:231-5.
99. Eder AF, Kennedy JM, Dy BA, et al. Limiting and detecting bacterial contamination of apheresis platelets: Inlet-line diversion and increased culture volume improve component safety. *Transfusion* 2009;49:1554-63.
100. Jenkins C, Ramírez-Arcos S, Goldman M, et al. Bacterial contamination in platelets: Incremental improvements drive down but do not eliminate risk. *Transfusion* 2011;51:2555-65.
101. Dunbar NM, Kreuter JD, Marx-Wood CR, et al. Routine bacterial screening of apheresis platelets on Day 4 using a rapid test: A 4-year single-center experience. *Transfusion* 2013;53:2307-13.
102. White SK, Schmidt RL, Walker BS, et al. Bacterial contamination rate of platelet components by primary culture: a systematic review and meta analysis. *Transfusion* 2020;60:986-96.
103. Walker BS, White SK, Schmidt RL, et al. Residual bacterial detection rates after primary culture as determined by secondary culture and rapid testing in platelet components: A systematic review and meta-analysis. *Transfusion* 2020;60:2029-37.
104. van der Meer PF, Ypma PF, van Geloven N, et al. Hemostatic efficacy of pathogen-inactivated vs untreated platelets: a randomized controlled trial. *Blood* 2018;132:223-31.
105. Prowse CV. Component pathogen inactivation: A critical review. *Vox Sang* 2013;104:183-99.
106. Simons FE, Arduzzo LR, Bilò MB, et al. World allergy organization guidelines for the assessment and management of anaphylaxis. *World Allergy Organ J* 2011;4:13-37.
107. Tobian AAR, King KE, Ness PM. Transfusion premedications: A growing practice not based on evidence. *Transfusion* 2007;47:1089-96.
108. Sandler SG, Eder AF, Goldman M, et al. The entity of immunoglobulin A-related anaphylactic transfusion reactions is not evidence based. *Transfusion* 2015;55:199-204.
109. Vlaar APJ, Juffermans NP. Transfusion-related acute lung injury: A clinical review. *Lancet* 2013;382:984-94.
110. Semple JW, Rebetz J, Kapur R. Transfusion-associated circulatory overload and transfusion-related acute lung injury. *Blood* 2019;133:1840-53.
111. Goldman M, Webert KE, Arnold DM, et al. Proceedings of a consensus conference: Towards an understanding of TRALI. In: *Transfusion Medicine Reviews*. W.B. Saunders, 2005:2-31.
112. Ozier Y, Muller JY, Mertes PM, et al. Transfusion-related acute lung injury: Reports to the French Hemovigilance Network 2007 through 2008. *Transfusion* 2011;51:2102-10.
113. van Stein D, Beckers EA, Sintnicolaas K, et al. Transfusion-related acute lung injury reports in the Netherlands: An observational study. *Transfusion* 2010;50:213-20.
114. Toy P, Gajic O, Bacchetti P, et al. Transfusion-related acute lung injury: Incidence and risk factors. *Blood* 2012;119:1757-67.
115. Finlay HE, Cassorla L, Feiner J, et al. Designing and testing a computer-based screening system for transfusion-related acute lung injury. *Am J Clin Pathol* 2005;124:601-9.
116. Silliman CC. The two-event model of transfusion-related acute lung injury. *Crit Care Med* 2006;34:S124-31.
117. Middelburg RA, Van Der Bom JG. Transfusion-related acute lung injury not a two-hit, but a multicausal model. *Transfusion* 2015;55:953-60.
118. Vlaar APJ, Toy P, Fung M, et al. A consensus redefinition of transfusion-related acute lung injury. *Transfusion* 2019;59:2465-76.
119. Haemovigilance by JRCs 2015 [Internet]. [cited 2020 Dec 10]. Available online: [http://www.jrc.or.jp/mr/english/pdf/Haemovigilance2015\\_en\\_JRCs.pdf](http://www.jrc.or.jp/mr/english/pdf/Haemovigilance2015_en_JRCs.pdf)
120. Haemovigilance by JRCs 2016 [Internet]. [cited 2020 Dec 10]. Available online: [http://www.jrc.or.jp/mr/english/pdf/Haemovigilance2016\\_en.pdf](http://www.jrc.or.jp/mr/english/pdf/Haemovigilance2016_en.pdf)
121. The Japan Red Cross Society. Transfusion information 1907-168 [Internet]. 2019 [cited 2020 Dec 13]. Available online: [http://www.jrc.or.jp/mr/news/pdf/yuketsuj\\_1907\\_168.pdf](http://www.jrc.or.jp/mr/news/pdf/yuketsuj_1907_168.pdf)
122. Semple JW, Rebetz J, Kapur R. Transfusion-associated circulatory overload (TACO): Time to shed light on the pathophysiology. *ISBT Sci Ser* 2018;14:136-9.
123. Lieberman L, Maskens C, Cserti-Gazdewich C, et al. A retrospective review of patient factors, transfusion practices, and outcomes in patients with transfusion-associated circulatory overload. *Transfus Med Rev* 2013;27:206-12.
124. Andrzejewski C, Casey MA, Popovsky MA. How we view and approach transfusion-associated circulatory overload: Pathogenesis, diagnosis, management, mitigation, and prevention. *Transfusion* 2013;53:3037-47.
125. Alam A, Lin Y, Lima A, et al. The prevention of

- transfusion-associated circulatory overload. *Transfus Med Rev* 2013;27:105-12.
126. Wiersum-Osselton JC, Whitaker B, Grey S, et al. Revised international surveillance case definition of transfusion-associated circulatory overload: a classification agreement validation study. *Lancet Haematol* 2019;6:e350-8.
  127. Roubinian NH, Hendrickson JE, Triulzi DJ, et al. Incidence and clinical characteristics of transfusion-associated circulatory overload using an active surveillance algorithm. *Vox Sang* 2017;112:56-63.
  128. Rana R, Fernández-Pérez ER, Khan SA, et al. Transfusion-related acute lung injury and pulmonary edema in critically ill patients: A retrospective study. *Transfusion* 2006;46:1478-83.
  129. Popovsky MA. Transfusion and the lung: Circulatory overload and acute lung injury. *Vox Sang* 2004;87:62-5.
  130. Murphy EL, Kwaan N, Looney MR, et al. Risk factors and outcomes in transfusion-associated circulatory overload. *Am J Med* 2013;126:357.e29-38.
  131. Pendergrast J, Armani C, Cserti-Gazdewich C, et al. Can furosemide prevent transfusion-associated circulatory overload? Results of a pilot, double-blind, randomized controlled trial. *Transfusion* 2019;59:1997-2006.
  132. Khandelwal A, Lin Y, Cserti-Gazdewich C, et al. TACO-BEL-3: a feasibility study and a retrospective audit of diuretics for patients receiving blood transfusion at ten hospitals. *Vox Sang* 2021;116:434-439.
  133. Panch SR, Montemayor-Garcia C, Klein HG. Hemolytic Transfusion Reactions. *N Engl J Med* 2019;381:150-62.
  134. Fung MK, Downes KA, Shulman IA. Transfusion of platelets containing ABO-incompatible plasma: A survey of 3156 North American Laboratories. *Arch Pathol Lab Med* 2007;131:909-16.
  135. Berséus O, Boman K, Nessen SC, et al. Risks of hemolysis due to anti-A and anti-B caused by the transfusion of blood or blood components containing ABO-incompatible plasma. *Transfusion* 2013;53 Suppl 1:114S-123S.
  136. Lozano M, Hedde N, Williamson LM, et al. Practices associated with ABO-incompatible platelet transfusions: a BEST Collaborative international survey. *Transfusion* 2010;50:1743-8.
  137. Quillen K. Hemolysis from platelet transfusion: call to action for an underreported reaction. *Transfusion* 2012;52:2072-4.
  138. Josephson CD, Castillejo MI, Grima K, et al. ABO-mismatched platelet transfusions: Strategies to mitigate patient exposure to naturally occurring hemolytic antibodies. *Transfus Apher Sci* 2010;42:83-8.
  139. Kaufman RM, Assmann SF, Triulzi DJ, et al. Transfusion-related adverse events in the Platelet Dose study. *Transfusion* 2015;55:144-53.
  140. Shehata N, Tinmouth A, Naglie G, et al. ABO-identical versus nonidentical platelet transfusion: A systematic review. *Transfusion*. 2009;49:2442-53.
  141. Busch MP, Caglioti S, Robertson EF, et al. Screening the Blood Supply for West Nile Virus RNA by Nucleic Acid Amplification Testing. *N Engl J Med* 2005;353:460-7.
  142. Stramer SL, Fang CT, Foster GA, et al. West Nile Virus among Blood Donors in the United States, 2003 and 2004. *N Engl J Med* 2005;353:451-9.
  143. Gale SA, Williamson PC, Busch MP, et al. First Zika-positive donations in the continental United States. *Transfusion* 2017;57:762-9.
  144. Liu R, Wang X, Ma Y, et al. Prevalence of Zika virus in blood donations: a systematic review and meta-analysis. *BMC Infect Dis* 2019;19:590.
  145. Jimenez A, Shaz BH, Bloch EM. Zika Virus and the Blood Supply: What Do We Know? *Transfus Med Rev* 2017;31:1-10.
  146. Harvala H, Hewitt PE, Reynolds C, et al. Hepatitis E virus in blood donors in England, 2016 to 2017: from selective to universal screening. *Euro Surveill* 2019;24:1800386.
  147. Izopet J, Lhomme S, Chapuy-Regaud S, et al. HEV and transfusion-recipient risk. *Transfus Clin Biol* 2017;24:176-81.
  148. Hewitt PE, Ijaz S, Brailsford SR, et al. Hepatitis E virus in blood components: A prevalence and transmission study in southeast England. *Lancet* 2014;384:1766-73.
  149. Petrik J, Lozano M, Seed CR, et al. Hepatitis E. *Vox Sang* 2016;110:93-130.
  150. Hewitt PE, Davison K, Howell DR, et al. Human T-lymphotropic virus lookback in NHS Blood and Transplant (England) reveals the efficacy of leukoreduction. *Transfusion* 2013;53:2168-75.
  151. Dollard SC, Roback JD, Gunthel C, et al. Measurements of human herpesvirus 8 viral load in blood before and after leukoreduction filtration. *Transfusion* 2013;53:2164-7.
  152. Roback JD, Josephson CD. New insights for preventing transfusion-transmitted cytomegalovirus and other white blood cell-associated viral infections. *Transfusion*. 2013;53:2112-6.
  153. Rühl H, Bein G, Sachs UJH. Transfusion-Associated Graft-Versus-Host Disease. *Transfus Med Rev* 2009;23:62-71.
  154. Treleaven J, Gennery A, Marsh J, et al. Guidelines on the use of irradiated blood components prepared by the

- British Committee for Standards in Haematology blood transfusion task force. *Br J Haematol* 2011;152:35-51.
155. Fast LD. Developments in the prevention of transfusion-associated graft-versus-host disease. *Br J Haematol* 2012;158:563-8.
  156. Uchida S, Tadokoro K, Takahashi M, et al. Analysis of 66 patients definitive with transfusion-associated graft-versus-host disease and the effect of universal irradiation of blood. *Transfus Med* 2013;23:416-22.
  157. Asai T, Inaba S, Ohto H, et al. Guidelines for irradiation of blood and blood components to prevent post-transfusion graft-vs.-host disease in Japan. *Transfus Med* 2000;10:315-20.
  158. Marschner S, Fast LD, Baldwin WM, et al. White blood cell inactivation after treatment with riboflavin and ultraviolet light. *Transfusion* 2010;50:2489-98.
  159. Hawkins J, Aster RH, Curtis BR. Post-transfusion purpura: Current perspectives. *J Blood Med* 2019;10:405-15.
  160. Win N, Peterkin MA, Watson WH. The Therapeutic Value of HPA-1a-Negative Platelet Transfusion in Post-Transfusion Purpura Complicated by Life-Threatening Haemorrhage. *Vox Sang* 1995;69:138-9.

doi: 10.21037/aob-20-90

**Cite this article as:** Sato T, Tsuno NH, Goto N, Hagino T, Tasaki T. Incidence and severity of adverse effects related to platelet transfusion: a narrative review of the literature and the recent hemovigilance data of Japan. *Ann Blood* 2021;6:24.